

## Diversity of marine ascomycetes from Maharashtra (India)-III

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### Abstract

Present paper deals with four marine fungi: *Haiyanga salina*, *Lignincola laevis*, *Lulworthia grandispora*, *Verruculina enalia* that were isolated from intertidal wood and foam samples from the coast of Thane District of Maharashtra.

**Keywords:** foam samples, intertidal wood, marine fungi

### Introduction

The study of marine fungi continues to attract mycologist, ecologist, physiologist and those interested in bioactive compounds. In the last decades, research on natural products (bioactive compounds) of marine fungi is on the rise and there are over 1,000 bioactive compounds described from marine fungi. From marine habitats, majority of the fungi recorded so far were Ascomycetous forms. Jones *et al.* (2015) [10] classified this group as 805 species in 352 genera. Using data from molecular studies, a number of new families have been added to the list of marine taxa within the Dothideomycetes (Kohlmeyer *et al.* 1995; Inderbitzin *et al.* 2001; Mugambi and Huhndorf, 2009; Suetrong *et al.* 2009, 2011; Zhang *et al.* 2009a, b) [11, 9, 12, 28, 29, 32].

### Material and Methods

Samples of intertidal woody debris, drift wood, dead stems, roots, leaves and fruits of mangroves, and foam samples from sandy beaches were collected during 2012-2013 at low tide at study sites along the coastal regions of Thane district and placed in polythene bags. Samples were transported to the laboratory. Collections contaminated by sediments or fouling organisms were washed with seawater. Specimens were observed for sporulating structures (Ascomata). After initial observations, samples were placed in plastic boxes and after two weeks examined for the presence of fungal fruiting bodies. Permanent voucher slides were made by using double cover glass method (Volkmann-Kohlmeyer and Kohlmeyer, 1996). Identifications of marine fungi were confirmed with the help of Monographs and illustrated taxonomic keys provided by Kohlmeyer and Kohlmeyer (1979), Kohlmeyer and Volkmann - Kohlmeyer (1991), Hyde *et al.* (2000) [27], Jones *et al.* (2009) [29], Borse *et al.* (2012) [3] and other relevant published literature.

### Taxonomic account

1. *Haiyanga salina* (Meyers) K.L Pang & E.B.G. Jones  
The Raffles Bull. Zool., 19- 8 (2008). = *Arenariomyces*

*salina* Meyers, *Mycologia*, 49-505 (1957); = *Remispora salina* (Meyers) Kohlm., *Mycologia*, 60-262 (1968); = *Halosphaeria salina* (Meyers) Kohlm., *Can. J. Bot.*, 50-1957 (1972); = *Antennospora salina* (Meyers) Yusoff *et al.* *Mycol. Res.*, 98-1003 (1994).

*Ascomata* are 100-425  $\mu\text{m}$  in diam. globose or subcylindrical, completely or partly immersed or becoming exposed, ostiolate, papillate, membranaceous or coriaceous, dark brown or black, surrounded by brown hyphae, solitary or gregarious. *Peridium* is 8-12  $\mu\text{m}$  thick. *Necks* are 68-165  $\mu\text{m}$  long, 28-52  $\mu\text{m}$  in diam. *Catenophyses* present. *Asci* were not seen as deliquising early. *Ascospores* are 19-28 x 8-13.5  $\mu\text{m}$  (excluding appendages), ellipsoidal, 1-septate, not or slightly constricted at the septum, hyaline, appendaged. *Appendages* at each end 3 or 4 (rarely five) radiating appendages, 12-19  $\mu\text{m}$  long, 1.5-2.5  $\mu\text{m}$  in diam at the base, subterminal, obclavate, curved, attenuate, semirigid, indistinctly spoon shaped at the base and release a drop of mucilage from their tips.

### Material examined

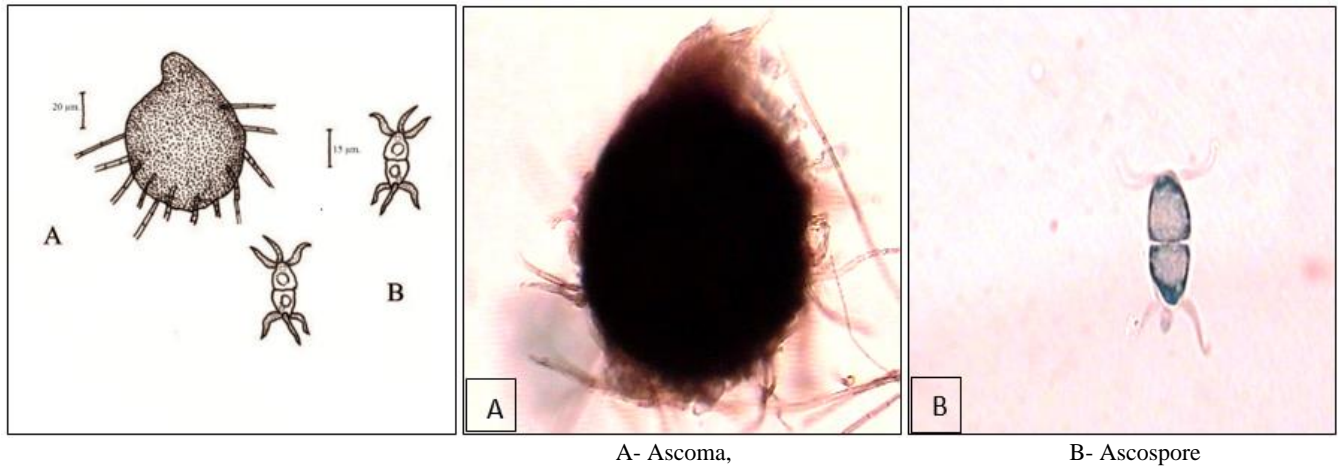
On intertidal wood, Dahanu; S. A. Gosavi 1112 (PGDB), 02 April 2011.

### Distribution in India-West Coast

Diu Island- Borse *et al.* (1999b) [7]; Guj.- Patil and Borse (2001) [18]; M.S.- Borse (1987a) [1]; Goa- Borse *et al.* (1999a) [5]; Kerala- Raveendran and Manimohan (2007) [14]; East Coast- T.N.- Nambiar and Raveendran (2009k) [17]; W.B.- Pawar and Borse (2004) [21].

### Remark

The descriptions and measurements of *Ascomata*, *Asci* and *Ascospores* are agreed with that of *H. salina* (Meyers) K.L Pang and E.B.G. Jones (2008). Therefore, it is assigned to that species. It is being reported for the first time from Thane district.



A- Ascoma,

B- Ascospore

Fig 1

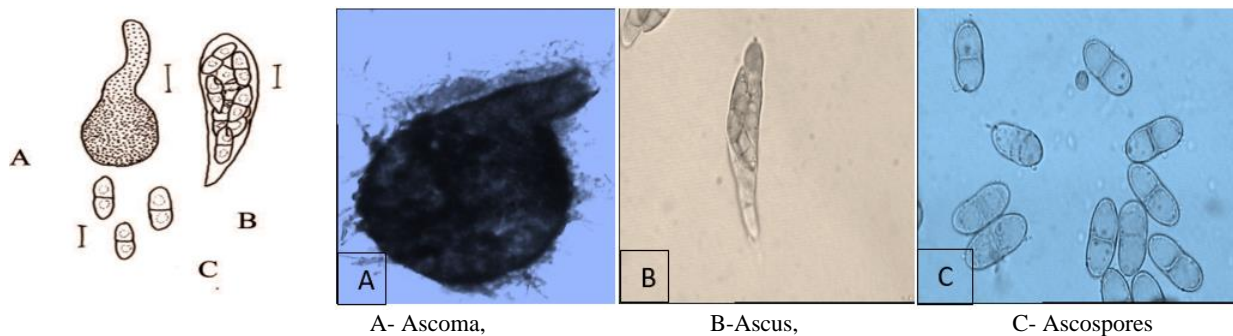
2. *Lignicola laevis* Hohnk Veroeff. *Inst. Meeresforsch. Bremerhaven*, 3- 216 (1955).

*Ascomata* are 132-254  $\mu\text{m}$  in diam., subglobose or ellipsoidal, immersed or superficial, ostiolate, papillate, coriaceous, hyaline, light brown, fuscous or blackish, solitary or gregarious. *Peridium* composed of 2-5 layers of elongate thick-walled cells with large lumina, forming a *textura angularis*. *Necks* are upto 4 mm long and 28-40  $\mu\text{m}$  in diameter. *Catenophyses* are of pseudoparenchyma of thin-walled cells with large lumina filling the venter of young ascomata, eventually breaking up into catenophyses. *Ascus* 46-68 x 14-22  $\mu\text{m}$ . 8-spored, clavate or subfusiform, short pedunculate, unitunicate, thin-walled, aphysoclastic, without apical apparatuses, persistent; *Ascospores* are 14-24 x 6-8  $\mu\text{m}$ , irregularly biseriate, ellipsoidal, one-septate, slightly constricted at the septum, hyaline, without appendages.

**Material examined:** On intertidal wood of *Avicennia officinalis*, Kalamb-Rajodi; S. A. Gosavi 1113 (PGDB), 8 July 2013.

**Distribution in India- East coast-** T.N.- Raghu kumar (1973) [13]; A.P.- Vittal and Sarma (2005) [30]; W.B.- Pawar and Borse (2004) [21]; *West coast-* M.S.- Patil and Borse (1983a); Goa- Borse *et al.* (1999a); Kerala- Nambiar and Raveendran (2006); Daman- Borse *et al.* (2000b); Guj.- Patil and Borse (2001) [19]; P.M.- Nambiar and Raveendran (2008d) [16].

**Remarks:** The descriptions and measurements of *Ascomata*, *Asci* and *Ascospores* are agreed with that of *L. laevis* Hohnk (Jones *et al.* 2009) [29]. Hence, it is assigned to that species. It is being reported for the first time from Thane district.



A- Ascoma,

B-Ascus,

C- Ascospores

Fig 2

3. *Lulworthia grandispora* Meyers, *Mycologia*, 49-513 (1957). = *L. grandispora* var. *apiculata* T.W. Johnson, *Mycologia*, 50-159 (1958).

*Ascomata* are 188-320  $\mu\text{m}$  in diam. globose or subglobose to pyriform, immersed or superficial, ostiolate, with long neck, brown to black, solitary or gregarious. *Necks* are 75-1400 x 15-33  $\mu\text{m}$ , cylindrical, straight or curved, sometimes two on one ascomata. *Asci* are 490-524 x 22-28  $\mu\text{m}$ . 8-spored, elongate-fusiform or cylindrical, unitunicate, thin-walled, early deliquescing. *Ascospores* are 508-780 x 2.5-5  $\mu\text{m}$  (including apical chambers), filamentous, curved, hyaline, tapering at each end into an elongate conical process or apical chamber. *Apical Processes* are (3-7  $\mu\text{m}$ .) long, acute

or rounded filled with mucus that is released through an apical pore.

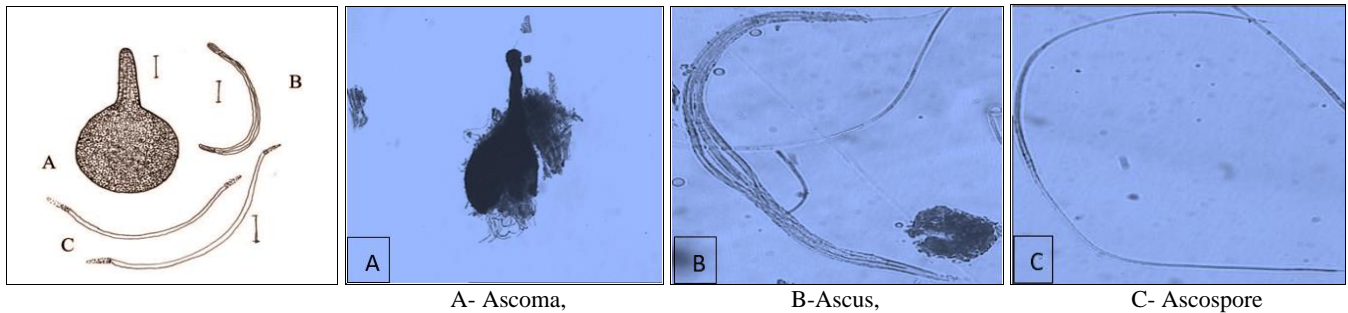
**Material examined:** On intertidal wood, Ghodbundar; S. A. Gosavi 1114 (PGDB), 20 Dec. 2013.

**Distribution in India-East coast:** T.N.- Nambiar and Raveendran (2006) [13]; A.P.- Sarma and Vittal (2000) [27]; W.B.- Pawar and Borse (2004) [21]; *West coast--* M.S.- Ramesh and Borse (1989) [24]; Goa- Tuwar (2009); Kerala- Nambiar and Raveendran (2006) [13]; Diu- Borse *et al.* (1999b) [7]; Daman- Borse *et al.* (2000b) [6]; Guj.- Borse *et al.* (2000a); P.M.- Nambiar and Ravindran (2007). (Source- Borse *et al.* 2012, 2013) [2, 3].

**Remarks**

The descriptions and measurements of Ascomata, Asci and Ascospores are completely agreed with that of *Lulworthia*

*grandispora* Meyers (Jones *et al.* 2009) [29]. Therefore, it is assigned to that species. It is being reported for the first time from Thane district.

**Fig 3**

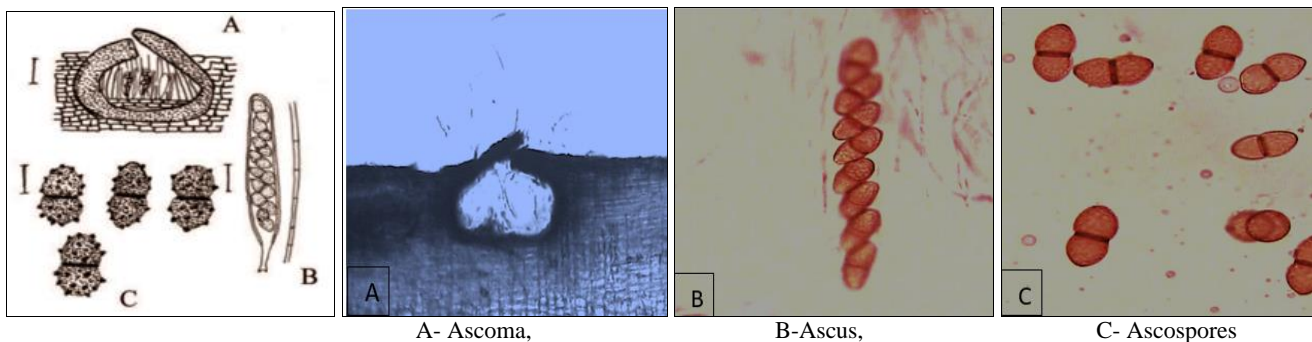
4. *Verruculina enalia* (Kohlm.) Kohlm. & Volkm.-Kohlm. *Mycol. Res.* 94: 689 (1990).  
= *Didymosphaeria enalia* Kohlm, *Ber. Deutsch. Bot. Ges.*, 79: 28 (1966).

*Ascomata* are 286–494  $\mu\text{m}$  high, 265–474  $\mu\text{m}$  in diam. subglobose, ampulliform or depressed ellipsoidal, partly or completely immersed, ostiolate, papillate, clypeate, carbonaceous, black and solitary. *Peridium* is 8–14  $\mu\text{m}$  thick. *Necks* are 72–144  $\mu\text{m}$  long, 135–310  $\mu\text{m}$  in diameter (including clypeus). *Pseudoparaphyses* are 1.5–2  $\mu\text{m}$  in diameter, septate, rarely branched, attached at both ends and reaching into the ostiolar canal; connected to each other by a gelatinous outer layer. *Asci* are 118–134 x 11–14  $\mu\text{m}$ . 8-spored, cylindrical, pedunculate, bitunicate, thick-walled, physoclastic, without apical apparatuses are developing at the base of the ascomata venter. *Ascospores* are 16–24 x 6.5–10  $\mu\text{m}$ , obliquely uniseriate, ellipsoidal, one-septate,

constricted at the septum, dark brown, verrucose to verruculose.

**Material examined:** On intertidal wood of *Avicennia marina*, Arnala; S. A. Gosavi 1116 (PGDB), 15 April 2013.

**Distribution in India-East coast:** T.N- Nambiar *et al.* (2008b) [15]; Pondicherry Karaikkal (P.K.)- Ravi kumar *et al.* (2009) [29]; A.P.- Sarma and Vittal (1998-99); W.B.- Pawar *et al.* (2009) [22]; Orissa- Borse *et al.* (2009) [8]; A. & N. Islands- Chinnaraj (1993a). *West coast-* M.S.- Patil and Borse (1985) [20]; Kerala- Nambiar and Raveendran (2006) [13]; Diu- Borse *et al.* (1999b) [7]; Goa- Borse *et al.* (1999a) [5]; Daman- Borse *et al.* (2000b) [6]; Guj.- Borse *et al.* (2000a) [4]; P.M.- Nambiar and Raveendran (2008d) [16]; Goa- Borse *et al.* (1999a) [5] (Source- Borse *et al.* 2012, 2013) [2, 3].

**Fig 4****Acknowledgement**

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**References**

1. Borse BD. Marine fungi from India - IV. *Trans. Mycol. Soc. Japan.* 1987a; 28:55-61.
2. Borse BD, Borse KN, Pawar NS, Tuwar AR. Marine fungi from India – XII. A Revised check list. *Ind. Jour. Geo-Mar. Sci.* 42:110-119. Fungi from Pirotan Island (Gujarat), India. *Geobios.* 2013; 27:145-148.
3. Borse BD, Borse KN, Pawar NS, Tuwar AR. “*Marine*

*Fungi of India (Monograph)*”, Broadway Book Centre Publishers and Distributors, Panjim, Goa, 2012, 1-471.

4. Borse BD, Kelkar DJ, Patil AC. Frequency of occurrence of marine, 2000a.
5. Borse BD, Nandan SN, Shinde DN. Higher marine fungi from Goa Coast (India). *J. Adv. Sci. Tech.* 1999a; 2:52-55.
6. Borse BD, Patil KB, Patil RV, Kelkar DJ. Marine fungi in foam, intertidal wood and dead *Avicennia marina* wood from Daman Coast, India. *Geobios.* 2000b; 27:42-44.
7. Borse BD, Patil RV, Kelkar DJ. Marine fungi from Div Island (India). *J. Adv. Sci. Tech.* 1999b; 11:1-8.
8. Borse KN, Pawar NS, Borse BD. Marine fungi from Orissa (India) - VI. In- *Biodiversity, Sustainable*



- Development and Human Welfare, (Eds. Nandan *et al.*), Pub. SSVPS's Sci.College, Dhule, Maharashtra. 2009; 35-41.
9. Inderbitzin P, Landvic S, Abdel-Wahab MA, Berbee ML. Aliquandostipitaceae, a new family for two new tropical ascomycetes with unusually wide hyphae and dimorphic ascomata. *Amer. J. Bot.* 2001; 88:52-61.
  10. Jones EBG, Suetrong S, Sakayaroj J, Bhalkali AH, Abdel-Wahab MA, Boekhout Teun, et al. Classification of marine Ascomycota, Basidiomycota, Blastocladiomycota and Chytridiomycota. *Fungal Diversity*- 2015; 73:1-72.
  11. Kohlmeyer J, Volkmann-Kohlmeyer B, Eriksson OE. Fungi on *Juncusroemerianus*. 2. New dictyosporous ascomycetes. *Bot. Mar.* 1995; 38:165-174.
  12. Mugambi GK, Huhndorf SM. Molecular phylogenetics of Pleosporales- Melanommataceae and Lophiostomataceae re-circumscribed (Pleosporomycetidae, Dothideomycetes and Ascomycota). *Studies Mycology.* 2009; 64:103-121.
  13. Nambiar G, Raveendran KA. Comparative account of Pokkali and mangrove associated marine and manglicolous marine fungi from Valapattanam estuary, Kannur District (Kerala). *Extended Abstract*, 18<sup>th</sup>. *Kerala Science Congress*, CESS, Akkulam, Thiruvananthapuram, 2006, 559-561.
  14. Nambiar GR, Raveendran K. Estuary's marine mycoflora of North Malabar (Kerala). *J. Mar. Atmos. Res.* 2007; 3:29-31.
  15. Nambiar GR, Raveendran K. Marine and manglicolous fungal diversity in the coastal wetlands of Kerala. *Seaweed Res. Utiln., (Special Issue).* 2008b; 30:107-111.
  16. Nambiar G, Raveendran K. Impact of coir retting on manglicolous marine fungi of Kerala coastal waters. *Poll. Res.* 2008d; 27:481-483.
  17. Nambiar GR, Raveendran K. Marine mycoflora of south India with special emphasis to lignicolous marine fungi. *Front. Biol. China.* 2009k; 4:436-441.
  18. Patil KB, Borse BD. Studies on higher marine fungi from Gujarat coast (India). *Geobios.* 2001; 28:41-44.
  19. Patil SD, Borse BD. Marine fungi from Maharashtra (India) - II. Some Arenicolous fungi. *Indian Bot. Repr.* 1983a; 2:86-87.
  20. Patil SD, Borse BD. Marine fungi from Maharashtra (India) - IV- Some Loculoascomycetes. *Trans. Mycol. Soc. Japan.* 1985; 26:271-276.
  21. Pawar NS, Borse BD. Marine fungi from Sundarbans (India) -IV. *J. Adv. Sci. & Tech.* 2004; 7:17-28.
  22. Pawar NS, Borse KN, Borse BD. Marine fungi from Sundarbans (India) -VII. In- *Biodiversity, Sustainable Development and Human Welfare*, (Eds. Nandan *et al.*), Pub.SSVPS's Sci.College, Dhule, Maharashtra. 2009; 91-103.
  23. Raghu Kumar S. Marine lignicolous fungi from India. *Kavaka.* 1973; 1:73-85.
  24. Ramesh Ch, Borse BD. Marine fungi from Maharashtra Coast (India). *Acta Botanica Indica.* 1989; 17:143-146.
  25. Raveendran K, Manimohan P. Eds. "Marine Fungi of Kerala, A Preliminary Floristic and Ecological Study", Malbar Natural History Society, Calicut. Kerala, India. 2007; 1-270.
  26. Ravi kumar M, Sridhar KR, Siva kumar T, Karam chand KS, Sivakumar N, Vellaiyan R. *et al.* Diversity of filamentous fungi on coastal woody debris after tsunami on the southeast coast of India. *Czech. Mycol.* 2009; 61:107-115.
  27. Sarma VV, Vittal BPR. Biodiversity of mangrove fungi on different substrata of *Rhizophora apiculata* and *Avicennia sp.* from Godavari and Krishna deltas, East coast of India In - *Aquatic mycology across the Millenium* (Eds KD. Hyde, WH Ho, SB. Pointing), *Fungal Diversity.* 2000; 5:23-41.
  28. Suetrong S, Hyde KD, Zhang Y, Bahkali AH, Jones EBG. Trematosphaeriaceae fam. nov. (Dothideomycetes, Ascomycota). *Cryptog. Mycolg.* 2011; 32:343-358.
  29. Suetrong S, Schoch CL, Spatafora JW, Kohlmeyer J, Volkmann-Kohlmeyer B, Sakayaroj J, *et al.* Molecular systematics of the marine Dothideomycetes. *Stud. Mycol.* 2009; 64:155-173.
  30. Vittal BPR, Sarma VV. Fungal Diversity on mangroves. In- "The fungaldiversity and conservation in India" (eds. Dargan, J. S., Arti, N. S. & Dhingara, G. S.), Bishen Singh Mahendra Pal Singh, Dehra Dun India. 2005; 33-45.
  31. Zhang Y, Schoch CL, Fournier J, Crous PW, de Gruyter J, Woudenberg JHC, *et al.* Multi-locus phylogeny of Pleosporales- a taxonomic, ecological and evolutionary re-evaluation. *Studies Mycology.* 2009b; 64:85-103.
  32. Zhang Y, Wang HK, Crous PW, Pointing SB, Hyde KD. Towards a phylogenetic clarification of *Lophiostoma / Massarina* and morphological similar genera in the Pleosporales. *Fungal Diversity.* 2009a; 38:225-251.